



Dynamics of fibropapillomatosis in green turtles *Chelonia mydas* from Babitonga Bay, southern Brazil

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ABSTRACT: Green turtles *Chelonia mydas* are vital components of marine ecosystems and are recognized as indicators of environmental health. Fibropapillomatosis (FP), a debilitating disease associated with a chelonid herpesvirus, disproportionately affects juveniles in coastal foraging areas. This study presents the first analysis of FP dynamics in Babitonga Bay, southern Brazil, an important habitat for the species. Between 2019 and 2023, 171 juvenile green turtles were monitored using capture–mark–recapture and photo-identification techniques. FP prevalence in Babitonga Bay was among the highest reported in Brazil, reaching 53.2%. Tumors were predominantly located in the anterior region of the body, particularly around the flippers and axillary area, and were mostly small, consistent with mild FP. Disease severity was quantified using an FP index, and its association with body condition was examined, revealing no significant association. Data from recaptured individuals revealed both tumor progression and regression, underscoring individual variability and suggesting the influence of environmental and immunological factors. The predominance of FP in juveniles highlights their heightened vulnerability during early developmental stages. These findings enhance our understanding of FP dynamics in green turtles and support the need for continued health assessments and conservation measures. This research contributes to global efforts to safeguard marine biodiversity, aligning with the goals of the United Nations Decade of Ocean Science for Sustainable Development.

KEY WORDS: Sea turtle · Infectious disease · Fibropapilloma tumors · Epizootic · Atlantic Ocean

1. INTRODUCTION

Environmental degradation and pollution are major drivers of marine diseases, some of which have the potential to reach widespread epizootics (Herbst et al. 2004, Ene et al. 2005, Jones et al. 2016). Among these, fibropapillomatosis (FP) stands out as an epizootic neoplastic disease that predominantly affects green turtles *Chelonia mydas* (Linnaeus, 1758) (Testudines, Cheloniidae), especially juveniles in shallow coastal areas near human settlements (Herbst 1994, George 1996, Van Houtan et al. 2010, Jones et al. 2020, Kelley et al. 2022). By contrast, FP is rarely observed in adults and is virtually absent on remote nesting islands (Baptistotte 2016, Domiciano et al. 2017, Gattamorta et al. 2022).

Beyond its ecological role, the green turtle is considered an environmental sentinel, reflecting the health of marine ecosystems (Aguirre & Lutz 2004, Domiciano et al. 2017). This status supports the identification of critical areas and the implementation of measures to mitigate environmental impacts and improve animal health. The species is listed as Endangered by the IUCN Red List (Seminoff 2004), although the South Atlantic subpopulation is categorized as Least Concern (Broderick & Patricio 2019). In Brazil, it was recently removed from the Official National List of Threatened Fauna, where it had previously been listed as vulnerable (IBAMA 2014, 2022).

FP causes cutaneous tumors of varying size and location, affecting both external tissues and internal organs (Herbst & Klein 1995, Wyneken et al. 2013,

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Jones et al. 2016, Domiciano et al. 2017). These tumors can impair essential functions such as movement, feeding, and vision, and may be fatal in severe cases (Herbst 1994, Ene et al. 2005, Flint 2013). The global variability in FP presentation underscores the importance of region-specific studies (Balazs 1991, Adnyana et al. 1997, Lackovich et al. 1999, Aguirre et al. 2000, Espinoza-Rodríguez & Barrios-Garrido 2021, Monteiro et al. 2021, Roost et al. 2022). In Brazil, FP was first reported in 1986, and current prevalence rates range from 18.6 to 58% (Santos et al. 2010, Tagliolatto et al. 2016, Domiciano et al. 2019, Rossi et al. 2019, Silva-Júnior et al. 2019, Bastos et al. 2022, Miguel et al. 2022). Given its potential to compromise individual survival and population health, FP is a key concern for sea turtle conservation and remains a global research priority (Hamann et al. 2010, Rees et al. 2016).

Tumor development is associated with infection by scutavirus chelonidalpha5 (ChHV5) and is influenced by both environmental and immunological cofactors (Herbst et al. 2004, Ene et al. 2005, Jones et al. 2016, Zamana et al. 2021, Vanstreels et al. 2023). Despite advances in research, the mechanisms underlying tumor progression and regression remain poorly understood (Tagliolatto et al. 2016). This complexity is reflected in the wide range of clinical outcomes, from spontaneous tumor regression to severe and potentially fatal cases (Herbst 1994, Patrício et al. 2016, Kelley et al. 2022). Disease susceptibility is further modulated by host-related factors such as immunosuppression, genetic predisposition, and overall physiological condition (Aguirre 1991, Herbst 1994, George 1996, Farrell et al. 2021, Kelley et al. 2022).

Regional differences in FP expression may also be driven by the genetic diversity of the ChHV5. Phylogeographic analyses have identified 4 major viral genetic groups associated with distinct oceanic regions: central Pacific, eastern Pacific, western Atlantic–eastern Caribbean, and Atlantic (Herbst et al. 2004, Ene et al. 2005, Patrício et al. 2012). In Brazil, molecular studies in northern São Paulo detected 2 ChHV5 variants: one from the Atlantic group and another genetically similar to strains from Hawai'i and Taiwan, suggesting potential viral gene flow between geographically distant populations (Zamana et al. 2021). Such genetic variability may influence viral pathogenicity and contribute to the heterogeneous clinical presentation of FP across different regions.

In Babitonga Bay, an important developmental habitat for juvenile green turtles in southern Brazil, FP has been recorded since at least 2001 (Cremer & Sartori 2009, Cremer et al. 2020). The bay is also a strategic economic hub, supporting large-scale port opera-

tions and surrounded by municipalities with significant industrial activity (Cremer 2006). Understanding how FP manifests in this population and which biological parameters influence its prevalence and severity is essential for evaluating viral spread and population health. This study investigates FP in green turtles from Babitonga Bay by analyzing temporal prevalence, variation across size classes, associations between disease severity and body condition, and tumor progression in affected individuals.

2. MATERIALS AND METHODS

2.1. Study site

Babitonga Bay is located on the northern coast of Santa Catarina state, southern Brazil (26° 13' 51" S, 48° 38' 5" W) (Fig. 1). The estuary is characterized by an extensive mangrove area (approximately 160 km²) and an average depth of 6 m. It is connected to the Atlantic Ocean by a single channel approximately 1850 m wide. The bay is heavily impacted by domestic and industrial effluents resulting from nearby urban expansion and industrial development, and it also supports intense port activity (Cremer 2006, Gerhardinger et al. 2021). Despite these anthropogenic pressures, Babitonga Bay plays a key role in regional biodiversity and is recognized as a priority area for conservation (MMA/SBio/DCME 2023).

Sampling was carried out at 3 sites along the southern margin of Babitonga Bay, within the urban area of São Francisco do Sul municipality. All sites are affected by untreated domestic sewage, feature alternating sandy and rocky shores, and are exposed to low to moderate small-boat traffic. The sampling locations include Bela Vista (Point A), Figueira Beach (Point B), and Calixto Beach (Point C) (Fig. 1). Bela Vista is adjacent to the São Francisco do Sul Port, while the other 2 sites are progressively more distant from this area.

2.2. Intentional capture procedures

Intentional capture campaigns of green turtles were conducted between 2019 and 2023 using a 70 m long, 3 m high gillnet with a 25–30 cm mesh size, made of 0.60 mm monofilament nylon, set parallel to the rocky shore (Ehrhart & Ogren 1999). Continuous monitoring by 2 teams, one in the water and another in a small boat, ensured that entangled animals were promptly released to prevent drowning. Some individuals were

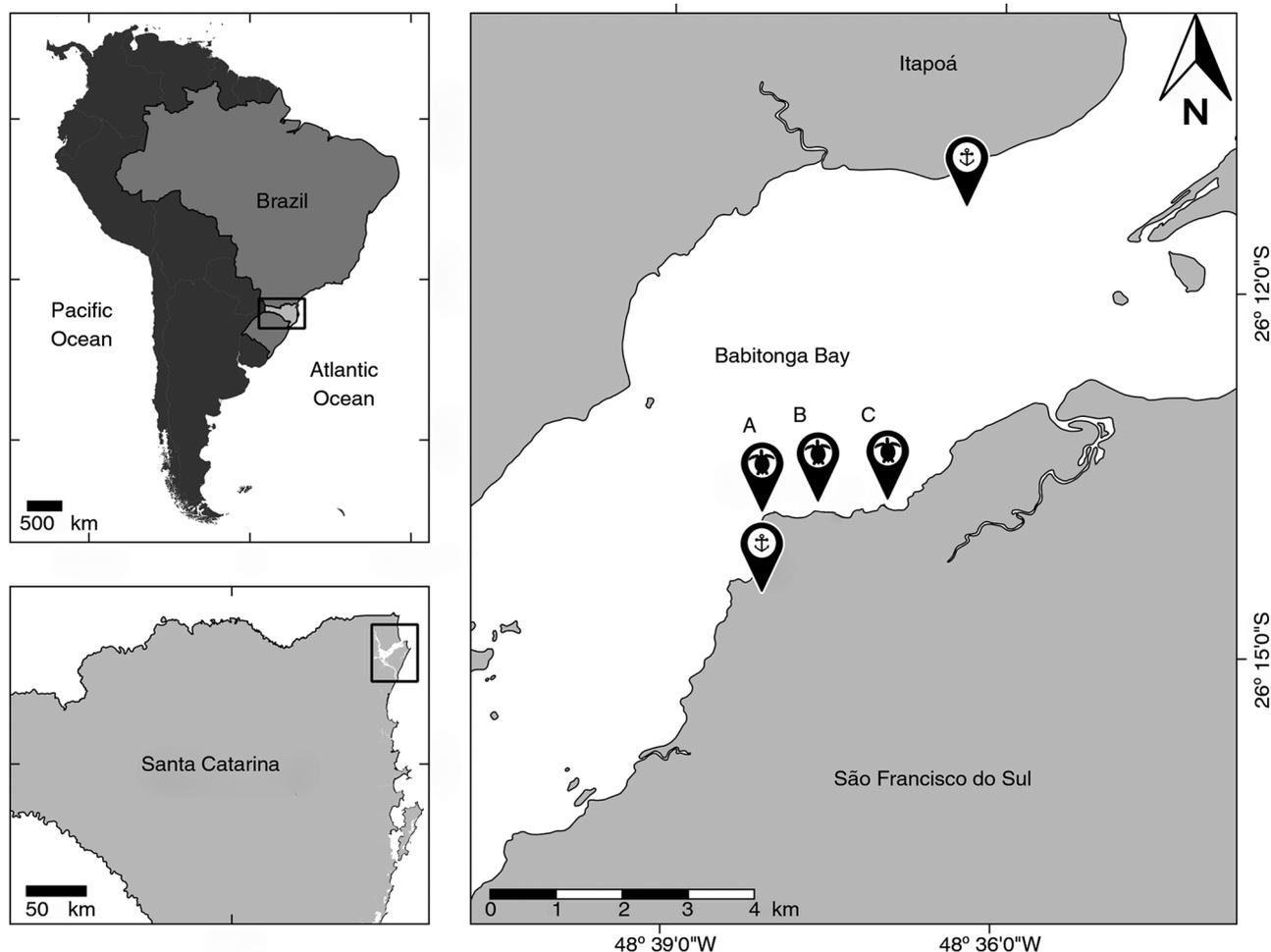


Fig. 1. Sampling locations of green turtles in Babitonga Bay, southern Brazil. Circles: capture sites (A, B, and C); anchor symbol: the port area

also captured manually. Turtles were restrained and transported to a land-based facility for data collection. Animals with FP were handled separately from FP-free animals. Gloves were changed between each animal, and 2 sets of sampling kits were used: one dedicated to FP turtles and another to turtles without FP. Disposable materials (e.g. scalpel blades, needles) were replaced for each animal, while reusable instruments (e.g. scissors, forceps, scalpel handles, calipers, and measuring tapes) were disinfected or sterilized between animals to avoid cross-contamination.

This research was conducted under federal authorization from the Biodiversity Authorization and Information System (SISBIO, permit no. 63333).

2.3. Data collection

For each green turtle, curved carapace length (CCL, cm) and body mass (in kg) were recorded using a flex-

ible measuring tape and a suspended mechanical scale, respectively (Bolten 1999). External tumors were measured with calipers and classified by their maximum diameter into 4 categories: A (<1 cm), B (1–4 cm), C (>4–10 cm), and D (>10 cm) (Work & Balazs 1999).

Tumor location was recorded as either ventral or dorsal, and each tumor was also classified by specific anatomical region: anterior (head, neck, front flippers), posterior (rear flippers, tail, inguinal), ocular (eyelid, periorcular, cornea), carapace, and plastron (Rossi et al. 2016, Tagliolatto et al. 2016). Each turtle was identified with 2 Inconel metal tags provided by the Sea Turtle Center of the Chico Mendes Institute for Biodiversity Conservation (Centro TAMAR/ICMBio), following standard tagging protocols (Centro TAMAR/ICMBio 2017). Individuals were photographed dorsally and ventrally to document marks and deformities on the body, and laterally on the head to record the head scale configuration, which is unique to each turtle and is used as a supplementary identification

method, a widely recognized non-invasive technique in sea turtle research (Bennett et al. 2000, Carpentier et al. 2016, Hancock et al. 2023, Neves-Ferreira et al. 2023).

2.4. Data analysis

Analyses were conducted using different data sets depending on the purpose: tumor macroscopic assessment used all available tumor records without repeating individuals; FP prevalence, severity analyses, and body condition used only the last capture of each individual; and annual prevalence and size-class analyses used the last capture per individual per year. Progression and regression of tumors, as well as the transition matrix, used all capture events for turtles with multiple recaptures.

Total FP prevalence for the study period was calculated by dividing the number of turtles with tumors by the total number of individuals captured. Annual variation in FP prevalence was assessed using Pearson's chi-squared test, and variation in prevalence across size classes (defined in 5 cm CCL intervals) was assessed using Fisher's exact test with simulated *p*-values (based on 10 000 replicates).

To examine temporal changes in turtle size, variation in mean CCL across years was analyzed using the non-parametric Kruskal-Wallis test, followed by Dunn's post hoc test. This was done to explore whether years with differences in turtle size coincided with variations in FP prevalence, without directly testing FP as a function of size. Differences in mean CCL between turtles with and without FP were assessed using the Mann-Whitney-Wilcoxon test to determine whether disease occurrence was related to body size. The relationship between CCL and average tumor size score was assessed using Spearman's rank correlation and linear regression to evaluate whether tumor size was related to body size.

To examine the relationship between FP occurrence (presence–absence) and turtle body size, binomial generalized linear mixed models (GLMMs) were fitted. Candidate models included CCL (cm) as a linear and quadratic predictor. Individual turtle ID was included as a random intercept to account for repeated captures of the same animal. Model selection was based on Akaike's information criterion (AIC), and the improvement of the quadratic term was assessed by a likelihood ratio test. Model fit was further quantified by marginal and conditional R^2 values. Predicted probabilities and 95% confidence intervals were generated from the final model and plotted against CCL.

Disease severity was assessed using the Fibropapillomatosis Index (FPI), calculated as $FPI = 0.1 \times nA + 1 \times nB + 20 \times nC + 40 \times nD$, where nA , nB , nC , and nD represent the number of tumors in size classes A, B, C, and D, respectively, found on each individual turtle. This index incorporates both the number and size of tumors to provide a weighted severity score. FPI values were classified according to the Southwestern Atlantic Fibropapillomatosis Severity Score (FPS_{SWA}) as follows: no FP ($FPI = 0$), mild ($0 < FPI < 40$), moderate ($40 \leq FPI < 120$), and severe ($FPI \geq 120$) (Rossi et al. 2016). Differences in mean turtle size among FPS_{SWA} categories were analyzed using the Kruskal-Wallis test.

The body condition index (BCI) was calculated for each turtle as $BCI = \text{body mass} \div SCL^3 \times 10\,000$ (Bjorndal et al. 2000), where SCL is the straight carapace length estimated by the formula $SCL = 0.93 \times CCL - 0.28$ (Rossi et al. 2019). Differences in BCI between turtles affected and unaffected by FP were tested using a Student's *t*-test. Relationships between BCI and FP severity were assessed by comparing BCI across FPS_{SWA} categories using 1-way ANOVA and by evaluating the association between BCI and FPI scores through linear regression.

Photographic comparisons were used to assess tumor progression and regression in recaptured turtles with a history of FP. Only capture pairs with high-quality photographs, enabling reliable evaluation, were included. Progression was defined as the appearance of new tumors or an increase in size, and regression as their reduction or disappearance. To visualize disease dynamics, a transition matrix was constructed for all recaptured turtles, using consecutive FPS_{SWA} to track progression, regression, or stability across captures.

Data normality was assessed using the Shapiro-Wilk test, and homogeneity of variances was verified using Levene's test where applicable. Non-parametric alternatives were applied when assumptions for parametric tests were violated. All statistical analyses were performed at a significance level of $\alpha = 0.05$ in R (v.4.3.1) (R Core Team 2023), using the packages 'readxl', 'ggplot2', 'rstatix', 'RcmdrMisc', and their dependencies (Wickham 2016, Fox & Marquez 2023, Kassambara 2023, Wickham & Bryan 2023). To account for multiple comparisons and reduce the likelihood of false positives, *p*-values were adjusted using the Benjamini-Hochberg method to control the false discovery rate (FDR; Glickman et al. 2014) within biologically meaningful families of hypotheses (e.g. temporal comparisons, morphometric measurements, body condition indices). Both raw *p*-values and FDR-

adjusted p-values (q-values) are reported, with $q < 0.05$ considered significant.

3. RESULTS

3.1. FP prevalence

A total of 331 capture events resulted in the capture of 171 individual green turtles, with CCLs ranging from 29.5 to 71.2 cm (mean: 43.5 ± 9.2 cm). Turtles affected by FP ranged from 32.5 to 66.4 cm (42.5 ± 6.8 cm; $n = 80$), while unaffected individuals ranged from 29.5 to 71.2 cm (44.7 ± 11.3 cm; $n = 91$). Overall FP prevalence was 53.2% (91 out of 171). No significant difference in mean size was found between FP-affected and unaffected turtles (Mann-Whitney U -test: $W = 3580.5$, $p = 0.855$, $q = 0.855$). FP prevalence varied significantly across size classes (Fisher's exact test: $p = 0.001$, $q = 0.005$), peaking in the 40–45 cm CCL class, decreasing in larger classes, with a slight increase in the 65–70 cm class (Fig. 2).

The quadratic GLMM provided the best fit to the data (AIC = 383.0) compared to the linear model (AIC = 409.5), with a significant improvement in fit (likelihood ratio test: $\chi^2 = 28.5$, $df = 1$, $p < 0.001$). The probability of FP exhibited a non-linear relationship with CCL, increasing with body size up to a certain point and declining thereafter (Fig. 3). Both the linear (estimate = 1.33 ± 0.37 , $p < 0.001$) and quadratic terms (estimate = -0.015 ± 0.004 , $p < 0.001$) were strongly significant. The vertex of the quadratic curve occurred at approximately 45 cm CCL, indicating that FP prevalence peaked around this size class. The marginal R^2 indicated that fixed effects (CCL and CCL^2) explained 26% of the variation in FP occurrence, while the conditional R^2 (including the random effect of turtle ID) reached 73%, reflecting substantial individual heterogeneity.

FP prevalence also varied significantly over the study period ($\chi^2 = 15.291$, $df = 4$, $p = 0.004$, $q = 0.008$), with the highest values observed in 2019. Although mean turtle size showed significant variation across years (Kruskal-Wallis: $\chi^2 = 10.259$, $df = 4$, $p = 0.036$, $q =$

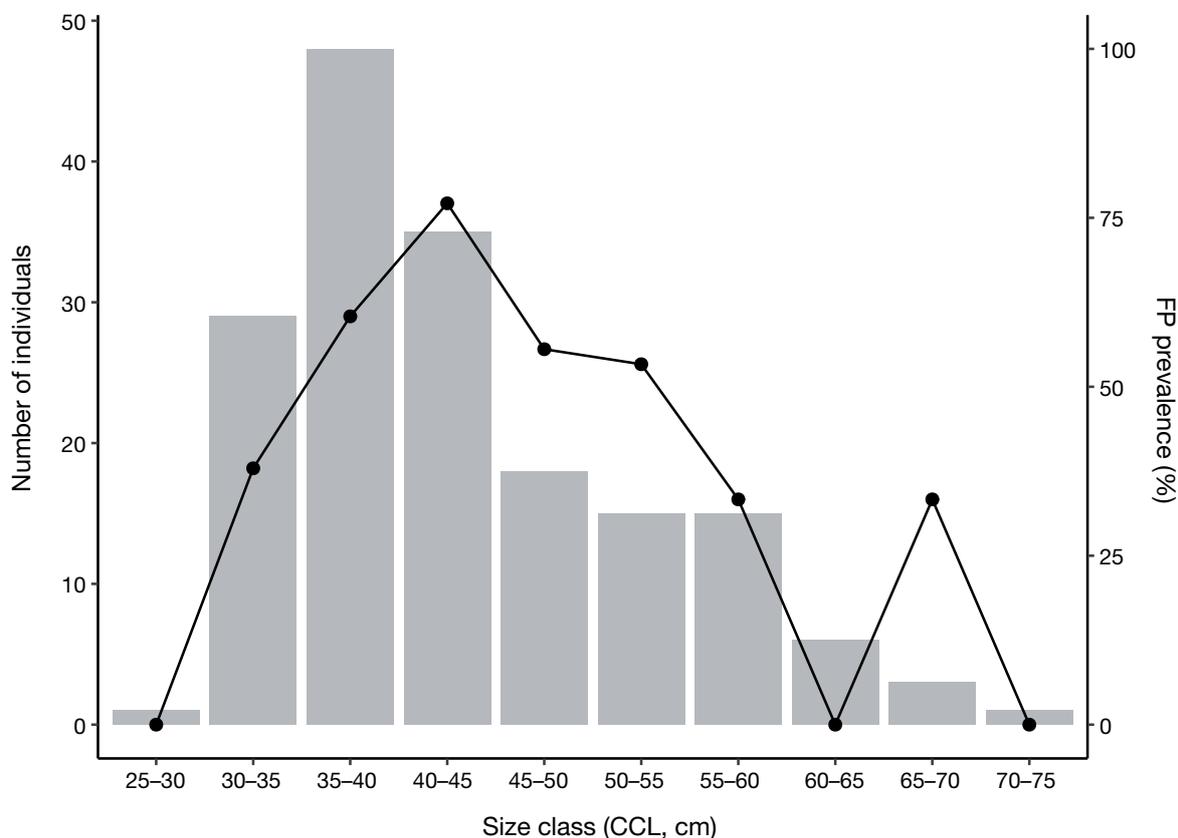


Fig. 2. Distribution of green turtle captures (left axis) and fibropapillomatosis (FP) prevalence (right axis) by curved carapace length (CCL) class in Babitonga Bay, southern Brazil. Prevalence values were scaled to match the range of the primary axis for visualization

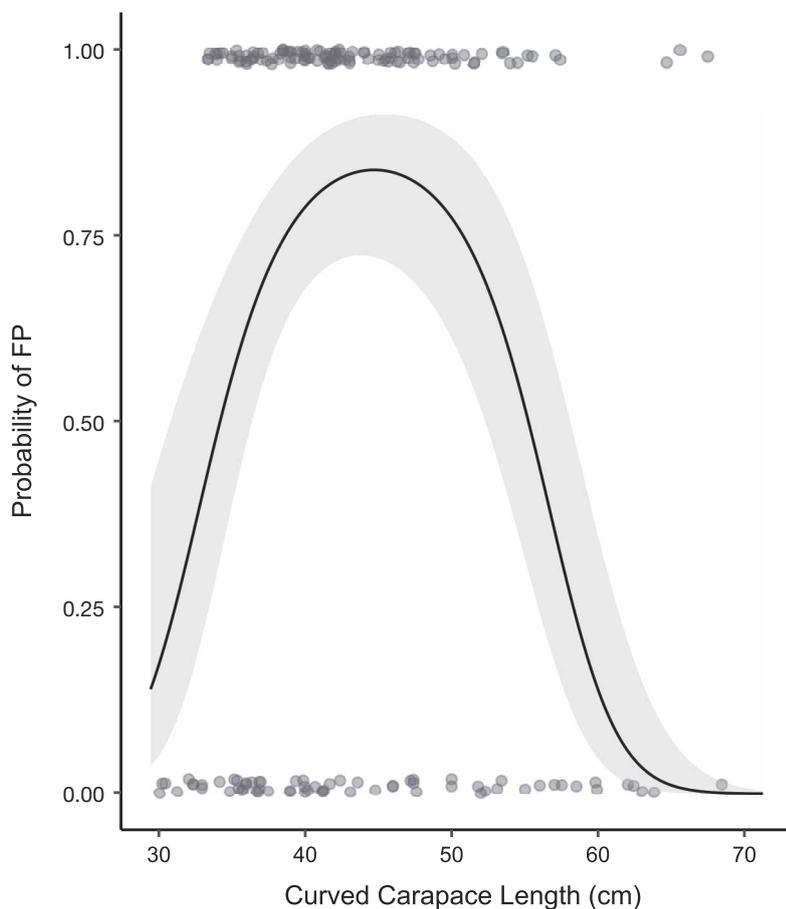


Fig. 3. Probability of fibropapillomatosis (FP) in green turtles from Babi-tonga Bay, southern Brazil, as a function of curved carapace length. Points: individual capture events (jittered for clarity); solid line: fitted quadratic generalized linear mixed model (with turtle ID as a random intercept); shaded area: 95% CI

0.036), post hoc pairwise comparisons using Dunn's test with Benjamini-Hochberg correction did not detect any statistically significant differences between individual years (all adjusted p -values > 0.05) (Fig. 4).

3.2. Macroscopic assessment of tumors

Tumors were assessed macroscopically using 98 available tumor records, each corresponding to a unique individual, regardless of recaptures. Lesions were observed in 95.9% of turtles in the ventral region and 52% in the dorsal region. The anterior region was affected in 84.7% of turtles, and the posterior region in 83.7%. Additionally, tumors were recorded on the carapace and/or plastron in 30.6% of individuals and in the ocular region in 23.5% (Table 1).

Of the 1459 tumors measured, 85.5% were located ventrally, 53.8% in the anterior region, and 38.7% in

the posterior. Carapace or plastron and ocular tumors accounted for 4.9 and 2.5%, respectively (Table 1).

While smaller tumors (Groups A and B) predominated and affected the majority of turtles, larger tumors (Group D) were rare and usually restricted to a single lesion per individual (Fig. 5). No correlation was found between CCL and tumor size score (Spearman's $\rho = 0.028$, $p = 0.610$, $q = 0.813$). Similarly, linear regression showed no significant relationship ($F_{1,329} = 1.324$, $r^2 = 0.004$, $p = 0.251$, $q = 0.502$).

3.3. FP severity and BCI

Of the 91 turtles that presented FP in their last capture, FPS_{SWA} were mild in 78%, moderate in 16.5%, and severe in 5.5%. FPI values ranged from 0.1 to 138.1 (mean: 26.1 ± 38.3). Mean CCL was 42.8 ± 7.27 cm in mild cases, 40.5 ± 4.00 cm in moderate, and 44.1 ± 6.59 cm in severe cases, with no significant differences among the severity levels (Kruskal-Wallis: $\chi^2 = 1.081$, $df = 3$, $p = 0.782$, $q = 0.855$).

BCI ranged from 1.14 to 2.06 in turtles without FP (1.56 ± 0.17) and from 1.14 to 1.96 in those with FP (1.58 ± 0.16), with no significant difference between groups (Student's t -test: $t = 1.2948$, $df = 154.77$, $p = 0.197$, $q = 0.592$) (Fig. 6A).

Similarly, BCI did not differ among turtles classified according to FPS_{SWA} : mild (1.58 ± 0.15 ; $n = 71$), moderate (1.58 ± 0.21 ; $n = 15$), and severe (1.50 ± 0.12 ; $n = 5$) (ANOVA: $F_{2,88} = 0.554$, $p = 0.577$, $q = 0.850$) (Fig. 6B). No significant correlation was observed between BCI and FPI (linear regression: $F_{1,82} = 0.03603$, $r^2 < 0.001$, $p = 0.850$, $q = 0.850$).

3.4. Progression and regression of FP

Tumor changes were assessed in 38 recaptured turtles with confirmed FP and suitable photographic records, yielding a total of 75 tumor comparison events. Based on individual turtles, tumor progression was observed in 17 individuals (44.7%), including 5 that developed tumors after initially showing no

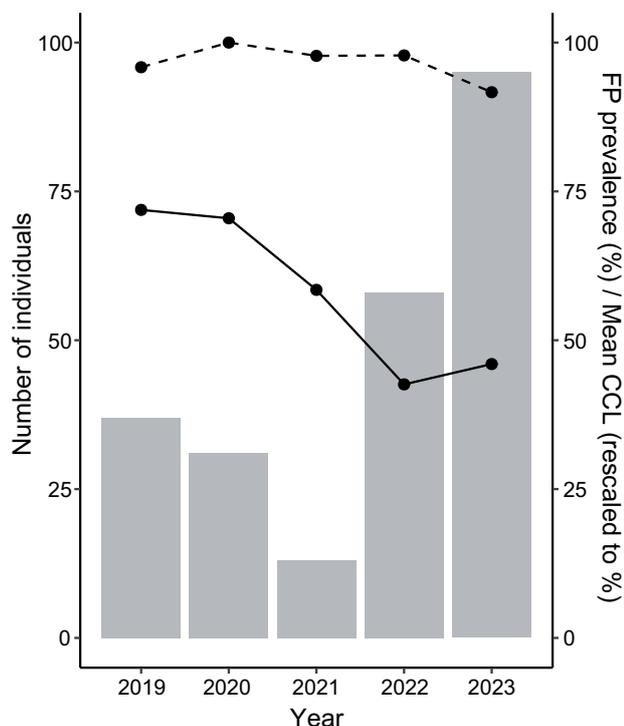


Fig. 4. Annual variation in FP prevalence (black line), mean curved carapace length (CCL; dashed line), and number of green turtles captured (bars) in Babitonga Bay, southern Brazil, from 2019 to 2023. Prevalence values were scaled to match the range of the primary axis, and mean CCL values were rescaled to a 0–100% range relative to the maximum observed mean CCL across all years (i.e. 100% corresponds to the year with the largest mean CCL). This rescaling allows visualization of trends in mean CCL alongside prevalence and capture numbers on a single plot

signs of FP (Fig. 7A). Tumor regression occurred in 14 individuals (36.8%) (Fig. 7B), with 7 individuals (18.4%) exhibiting both progression and regression between recaptures. An individual monitored over 4.5 yr (Fig. 8) showed tumor progression followed by partial regression.

At the event level, progression occurred in 41 events (54.7%), including 9 events with new tumor development, 14 with tumor growth, and 18 with both. Tumors appeared at an average (\pm SD) of 338 ± 285 d (range: 34–1116 d). Regression occurred in 34 events (45.3%), comprising 6 events of complete regression, 13 of partial regression, and 15 showing both. Tumors regressed at an average of 468 ± 392 d (range: 63–1242 d), whereas complete remission was observed after 793 ± 494 d (range: 188–1242 d).

A transition matrix based on FPS_{SWA} scores for all recaptured turtles (Table 2) revealed that most turtles initially classified as absent remained tumor-free (75%), with a few progressing to mild or moderate stages. Mild cases mostly remained stable (69%), while moderate cases either progressed to severe or regressed to mild, and severe cases were split between remaining severe or regressing to moderate.

4. DISCUSSION

The FP prevalence in Babitonga Bay (53.2%) ranks as the second highest value among Brazilian foraging grounds, only surpassed by Espírito Santo Bay (58.3%; Santos et al. 2010). Most other studies report lower prevalence values, generally between 18.6 and 43.1% (Tagliolatto et al. 2016, Rossi et al. 2019, Silva-Júnior et al. 2019, Bastos et al. 2022, Miguel et al. 2022, Vanstreels et al. 2023). Earlier records suggest that FP was historically rare or sporadic in the region: a study documented no FP cases among 30 stranded green turtles between 2004 and 2009, though 2 tumor-bearing turtles were reported in 2001 and 2002 (Cremer & Sartori 2009). The current high prevalence indicates a marked increase over the past 2 decades, although it remains below global FP hotspots such as Florida (72.5%; Lackovich et al. 1999), Australia (70%; Aguirre et al. 2000), Puerto Rico (up to 75%; Patrício et al. 2016), and Hawai'i (up to 92%; Balazs 1991), which has since declined to around 9% (Chaloupka et al. 2009).

As noted by Herbst (1994), FP prevalence is often positively correlated with proximity to agricultural, industrial, and urbanized areas. However, Monteiro et al. (2021) documented moderate prevalence in

Table 1. Anatomical distribution of tumors and number of affected green turtles in Babitonga Bay, southern Brazil. Regions: anterior (head excluding eyes, neck, and fore flippers); posterior (hind flippers, tail, and inguinal area); ocular (eyelid, periocular surfaces, and cornea)

	Tumors				Individuals	
	n	%	Min. –max.	Mean \pm SD	n	%
Body part						
Ventral	1247	85.5	1–49	13.3 ± 12.49	94	95.9
Dorsal	212	14.5	1–18	4.2 ± 3.43	51	52.0
Location on body						
Anterior region	784	53.8	1–39	8.0 ± 9.26	83	84.7
Posterior region	564	38.7	1–26	5.8 ± 6.25	82	83.7
Carapace and/or plastron	72	4.9	1–9	0.7 ± 1.65	30	30.6
Ocular region	37	2.5	1–3	0.4 ± 0.74	23	23.5
Total	1457 ^a	—	1–67	14.9 ± 14.68	98	—

^aTwo tumors had no recorded body location

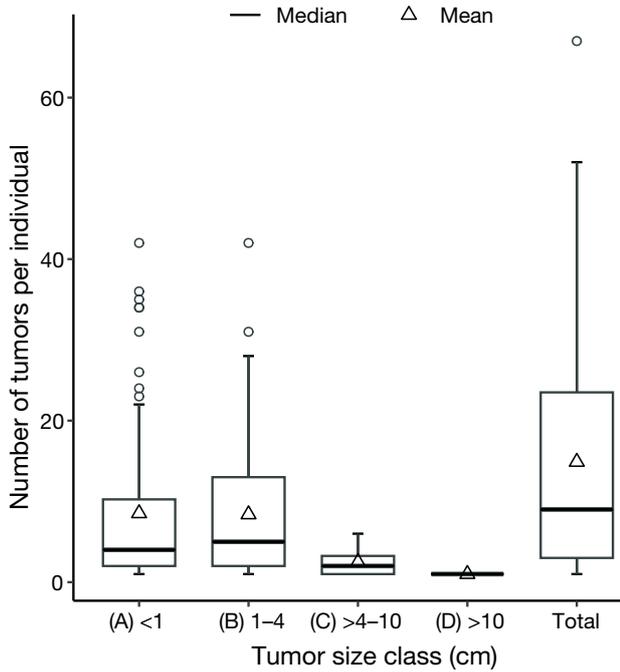


Fig. 5. Number of tumors per green turtle by tumor size class (A: <1 cm; B: 1–4 cm; C: >4–10 cm; D: >10 cm; n = 98) in Babitonga Bay, southern Brazil. Boxes: interquartile range (IQR; 25th–75th percentile); horizontal lines: median; whiskers extend to 1.5× the IQR; circles: outliers; triangles: mean values

minimally impacted habitats of West Africa, highlighting that FP expression may also depend on ecological and immunological factors beyond direct human disturbance. The elevated prevalence in Babitonga Bay may reflect regional environmental stressors such as chemical, microbiological, and noise pol-

lution (Santos et al. 2010). Additionally, a reduced diversity of algae in turtle diets may compromise nutritional intake and increase vulnerability to FP (Bjorndal 1996, Van Houtan et al. 2010). In Babitonga Bay, green turtles feed predominantly on chlorophytes, especially *Ulva*, with limited consumption of red and brown algae (Souza 2016), indicating low dietary diversity. Similar patterns, characterized by high FP prevalence and severity in areas where turtles consume low-diversity algal assemblages, have been reported elsewhere in Brazil (Bastos et al. 2022).

Importantly, even when turtles exhibit normal body condition, reduced diet diversity and altered chemical composition can still impair immune function, growth, and disease susceptibility, including FP (Bjorndal 1996, Van Houtan et al. 2010, Bastos et al. 2022). Maintaining high-quality feeding habitats with diverse algal communities is therefore essential for health, recruitment, and population resilience of green turtles (Worm et al. 2006, Rees et al. 2016, Mazaris et al. 2017).

All turtles in this study were classified as juveniles or subadults (CCL < 85 cm; Chaloupka & Limpus 2005, Lenz et al. 2017). FP prevalence was not evenly distributed across size classes, with the highest probability of occurrence in intermediate individuals and a peak around 40–45 cm CCL. Although sample sizes were uneven among size classes, with fewer individuals at the extremes of the size range, mean size did not differ significantly between affected and unaffected turtles. Similar ontogenetic patterns, with higher FP prevalence in intermediate juveniles fol-

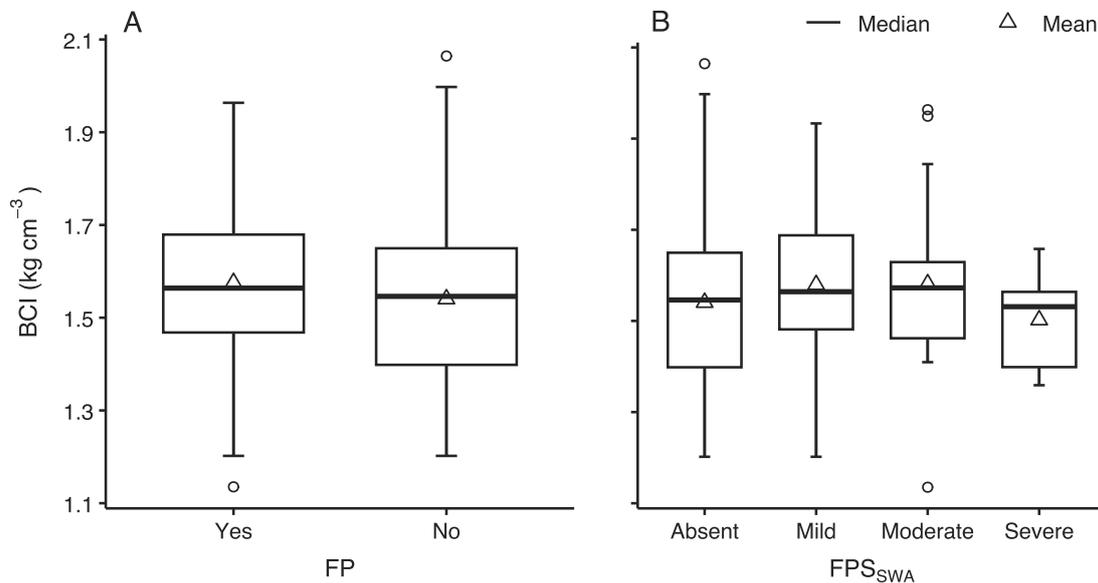


Fig. 6. (A) Body condition index (BCI) of green turtles with and without fibropapillomatosis (FP) in Babitonga Bay, southern Brazil. (B) BCI across Southwestern Atlantic Fibropapillomatosis Severity Score (FPS_{SWA}). Boxplot parameters as in Fig. 5



Fig. 7. Tumor progression and regression in 2 individual green turtles from Babitonga Bay, southern Brazil. (A) Progression in the inguinal region and hind flippers (714 d between captures). (B) Regression in the ocular region (red arrows) (832 d between captures)

lowed by reduced occurrence in larger turtles, have been reported in other Brazilian estuarine areas (Santos et al. 2010, Domiciano et al. 2019) and in West Africa (Monteiro et al. 2021), supporting the hypothesis of age-related resistance or a self-limiting disease course with tumor regression over time (Work et al. 2004, Foley et al. 2005, Patrício et al. 2016). The slightly earlier peak in Babitonga Bay compared to other regions may reflect local environmental conditions, such as estuarine productivity, water temperature, or habitat features (Herbst 1994, Zamana et al. 2021, Vanstreels et al. 2023).

FP is typically acquired after recruitment to coastal foraging areas (Herbst 1994, Ene et al. 2005). However, its detection in some newly recruited individuals (CCL < 35 cm; Lenz et al. 2017) underscores variability in recruitment sizes and exposure periods. While FP prevalence is generally concentrated in intermediate-size juveniles, these early cases are consistent with turtles that may have already spent close to 1 yr in coastal habitats, allowing sufficient time for infection. In our study, the single individual in the 25–30 cm size class showed no evidence of FP, reinforcing the expectation that newly recruited turtles typically arrive in nearshore waters without visible tumors. Tumor development in juveniles may also be influ-

enced by immunosuppression associated with ontogenetic shifts and local stressors (Santos et al. 2010, Domiciano et al. 2017), although current evidence indicates that immunosuppression is not a prerequisite (Cray et al. 2001, Work et al. 2001).

Molecular studies have shown that ChHV5 can be present in turtles without external tumors, including juveniles and apparently healthy adults (Quackenbush et al. 2001, Page-Karjian et al. 2012, Alfaro-Núñez et al. 2014), indicating that infection may occur before tumors develop. More recent environmental DNA (eDNA) approaches have even detected ChHV5 in sand associated with hatchling crawls, suggesting early exposure and the possibility of horizontal or vertical transmission prior to ocean entry (Farrell et al. 2022). In addition, a study on green turtle hatchlings in Florida found that all 297 sampled hatchlings tested negative for ChHV5 DNA, even when some adult females were positive, supporting the hypothesis that ChHV5 is not necessarily vertically transmitted, and that young turtles can hatch free of detectable infection (Page-Karjian et al. 2022). Together, these findings highlight that absence of visible tumors does not imply absence of infection and that subclinical or early-stage infections may be more common than previously recognized.

Turtles had tumors predominantly in specific body regions, with the highest frequency in the anterior (head, neck, front flippers) and posterior (rear flippers, tail, inguinal) regions. The larger surface area of soft tissue in the anterior region may account for this pattern, providing more suitable sites for tumor development (Baptistotte 2007). This anatomical distribution is commonly reported along the Brazilian coast, including Espírito Santo, Rio de Janeiro, São Paulo, and the northeastern region (Baptistotte 2007, Santos et al. 2010, Tagliolatto et al. 2016, Silva-Júnior et al. 2019, Miguel et al. 2022), as well as in Venezuela (Espinoza-Rodríguez & Barrios-Garrido 2021) and Hawai'i (Work et al. 2004). By contrast, an exception has been reported in Indonesia, where tumors predominantly occur in the posterior region (Adnyana et al. 1997), suggesting potential regional differences in tumor distribution. Tumors on the anterior flippers may impair swimming and foraging abilities, potentially reducing predator evasion and increasing mor-

ality. Turtles had tumors predominantly in specific body regions, with the highest frequency in the anterior (head, neck, front flippers) and posterior (rear flippers, tail, inguinal) regions. The larger surface area of soft tissue in the anterior region may account for this pattern, providing more suitable sites for tumor development (Baptistotte 2007). This anatomical distribution is commonly reported along the Brazilian coast, including Espírito Santo, Rio de Janeiro, São Paulo, and the northeastern region (Baptistotte 2007, Santos et al. 2010, Tagliolatto et al. 2016, Silva-Júnior et al. 2019, Miguel et al. 2022), as well as in Venezuela (Espinoza-Rodríguez & Barrios-Garrido 2021) and Hawai'i (Work et al. 2004). By contrast, an exception has been reported in Indonesia, where tumors predominantly occur in the posterior region (Adnyana et al. 1997), suggesting potential regional differences in tumor distribution. Tumors on the anterior flippers may impair swimming and foraging abilities, potentially reducing predator evasion and increasing mor-



Fig. 8. FP progression and regression in a green turtle recaptured 10 times in Babitonga Bay, southern Brazil, between 5 May 2019 and 11 December 2023. (A) First capture, showing no external tumors; (B) fifth recapture, 1001 d after first capture, with multiple new fibropapillomas; (C) ninth recapture, 680 d after fifth recapture, with partial tumor regression

Table 2. Transition matrix of fibropapillomatosis severity in recaptured green turtles (Southwestern Atlantic Fibropapillomatosis Severity Score, FPS_{SWA}) from Babitonga Bay, southern Brazil. Rows indicate the FPS_{SWA} score at the first capture, and columns indicate the score at the subsequent capture. Values represent the number of recapture events observed for each transition

FPS _{SWA}	Absent	Mild	Moderate	Severe
Absent	54	17	1	0
Mild	20	54	3	1
Moderate	0	2	3	3
Severe	0	1	0	1

bidity (Aguirre & Lutz 2004, Flint 2013, Jones et al. 2016). Ocular tumors were also observed, though infrequently and typically small (<1 cm), making visual impairment unlikely. This contrasts with historical reports from Hawai'i, where FP was observed to often affect the eyes and can lead to severe visual impairment (Work et al. 2004), highlighting possible geographic differences in tumor expression and severity.

Most tumors observed in our study were small (<1 cm), especially among individuals with CCL < 45 cm. This predominance of small tumors in smaller turtles suggests early-stage infections and is consistent with expectations for juveniles recently exposed to the virus. Although no significant relationship between turtle size and tumor size was found, similar patterns have been documented elsewhere and are often interpreted within the context of viral pathogenesis (Baptistotte 2007, Tagliolatto et al. 2016, Bastos et al. 2022, Miguel et al. 2022). Previous studies suggest that ChHV5 replication peaks during the early stages of infection (Rodenbusch et al. 2014, Work et al. 2015), contributing to the emergence of small tumors during the initial disease phase. As the disease progresses, viral replication may decrease, even as tumors continue to enlarge and persist (Work et al. 2004), supporting the hypothesis that FP follows an initial phase of intense viral activity followed by chronic tumor development.

According to FPS_{SWA} criteria, most turtles exhibited mild forms of the disease. This finding is compatible with reports from several feeding areas (Baptistotte 2007, Patrício et al. 2016, Tagliolatto et al. 2016, Rossi et al. 2019, Silva-Júnior et al. 2019, Monteiro et al. 2021, Vanstreels et al. 2023), while severe cases appear more frequent in Espírito Santo, Brazil (Santos et al. 2010, Miguel et al. 2022). Geographic differences in disease severity may reflect variation in local environmental conditions or in host-pathogen dynamics, including potential variation in the pathogenicity of

circulating ChHV5 strains (Ene et al. 2005, Zamana et al. 2021). While viral genotyping was not conducted in this study, genomic analyses of ChHV5 in other regions have demonstrated broad divergence between geographic strains and evidence of recombination, suggesting that viral variation may contribute to differences in disease manifestation (Morrison et al. 2018).

Although FP-affected turtles had slightly higher average BCI values than unaffected individuals, the difference was not statistically significant. This result is consistent with findings from other studies in Brazil (Tagliolatto et al. 2016, Santos et al. 2017, Rossi et al. 2019, Miguel et al. 2022) and elsewhere (Espinoza-Rodríguez & Barrios-Garrido 2021), which have reported either similar or unexpectedly higher BCI values in FP-affected turtles. This contrasts with earlier assumptions that FP would negatively affect body condition. In the Pacific, for example, FP has been associated with moderate to severe emaciation, cachexia, and other signs of poor condition in tumored turtles (Work et al. 2001), suggesting geographic variation in disease impacts. Several authors have suggested that the presence of numerous or large tumors can artificially inflate body mass, leading to elevated BCI values and potentially masking physiological impairment (Rossi et al. 2019, Vanstreels et al. 2023). In our case, most tumors were small, potentially minimizing this effect. Nevertheless, because BCI does not differentiate between muscle, fat, and abnormal tissue mass, it may lack the sensitivity needed to accurately reflect nutritional or health status in FP-affected individuals (Santos et al. 2015, Patrício et al. 2016, Domiciano et al. 2019, Vanstreels et al. 2023). This reinforces the need for caution in interpreting BCI values in the context of FP and suggests that future assessments might benefit from integrating tumor burden indices (e.g. FPI) to improve accuracy and diagnostic sensitivity.

Tumor progression occurred most frequently; however, we also documented tumor regression more often than previous studies reported in coastal areas such as Itaipu (Rio de Janeiro, Brazil; Machado Guimarães et al. 2013, Tagliolatto et al. 2016), Akumal Bay (Quintana Roo, Mexico; Muñoz Tenería et al. 2022), and Kwale County (Kenya; Hancock et al. 2023). This may reflect contextual differences in environmental or immunological factors affecting disease dynamics, such as local water quality, pollutant loads, or nutritional stressors that can influence immune competence and viral expression in sea turtles (Aguirre & Lutz 2004, Herbst et al. 2004, Bastos et al. 2022). Alternatively, regression in some individuals may indicate an intrinsic capacity to mount an effective immune

response capable of reducing or eliminating tumors over time. A similar case was recently described by Pérez et al. (2024), who documented complete regression of skin tumors in a juvenile green turtle in Itapirubá (Santa Catarina, southern Brazil) using photo-identification and histopathological analysis. These findings underscore the value of longitudinal monitoring, which enables the detection of such trends and provides insights into the timing and progression of FP lesions, patterns only discernible through repeated, long-term observations.

Stage transitions based on FPS_{SWA} scores indicated several events of turtles shifting from mild to absent categories. However, only 6 cases were classified as complete remission in the photographic analysis. This difference reflects that FPS_{SWA} transitions capture all recorded events, including repeated transitions for the same individual, whereas the photographic assessment was restricted to recaptures with high-quality images and to cases without subsequent relapse. Occasional inconsistencies in field records may also contribute. Therefore, the remission events reported here should be considered conservative estimates.

In our study, one individual (Fig. 6A) exhibited a skin marking in the femoral fossa region upon initial capture, possibly representing an early stage of tumor development. Similar markings are frequently observed in captured individuals; however, it remains unclear whether these represent incipient fibropapillomas or unrelated skin conditions, as this has not yet been formally investigated. Additionally, in another case (Fig. 8), a tumor developed at the site of a metal flipper tag. Although this was not examined in detail, field observations suggest that tumor development near tagging sites may be relatively common, highlighting the need for further research into possible associations between tagging procedures and the FP onset. We recorded a rare long-term FP trajectory in this individual, with tumors progressing and later regressing over intervals of several years, underscoring the potential for natural reversal of disease even after prolonged progression.

5. CONCLUSIONS

Patterns of FP observed in Babitonga Bay, including high prevalence among juveniles, predominance of small anterior tumors, and mostly mild clinical forms, are consistent with reports from other Brazilian coastal regions (Espírito Santo, Rio de Janeiro, São Paulo, and Paraná), suggesting common ecological drivers of FP across foraging grounds. Tumor progres-

sion was the most frequent outcome among recaptured individuals, although cases of regression, including complete remission, were also documented, indicating individual variation in disease trajectories.

These findings highlight the importance of long-term monitoring using capture–recapture data to track FP dynamics and inform conservation. Environmental stressors such as climate change, rising ocean temperatures, ongoing ecosystem degradation, and expansion of port activities may exacerbate FP outbreaks in the future, emphasizing the need to understand and mitigate both ecological and anthropogenic drivers. Continued monitoring of key foraging habitats like Babitonga Bay is essential to safeguard juvenile green turtles, refine population models, and support evidence-based management actions aimed at reducing disease impacts and promoting population resilience.

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LITERATURE CITED

- ✦ Adnyana W, Ladds PW, Blair D (1997) Observations of fibropapillomatosis in green turtles (*Chelonia mydas*) in Indonesia. *Aust Vet J* 75:737–742
- Aguirre AA (1991) Green turtle fibropapilloma: an epidemiologic perspective. In: Balazs GH, Pooley SG (eds) Research plan for marine turtle fibropapilloma. NOAA Tech Memo NMFS-SWFSC-156. US Department of Commerce, Honolulu, HI, p 107–113
- ✦ Aguirre AA, Lutz PL (2004) Marine turtles as sentinels of ecosystem health: Is fibropapillomatosis an indicator? *EcoHealth* 1:275–283
- Aguirre AA, Limpus CJ, Spraker TR, Balazs GH (2000) Survey of fibropapillomatosis and other potential diseases in marine turtles from Moreton Bay, Queensland, Australia. In: Kalb H, Wibbels T (eds) Proc 19th Ann Symp sea turtle biology and conservation, 2–6 March 1999, South Padre Island, TX. NOAA Tech Memo NMFS-SEFSC-443. National Marine Fisheries Service, Miami, FL, p 36
- ✦ Alfaro-Núñez A, Bertelsen MF, Bojesen AM, Rasmussen I, Zepeda-Mendoza L, Olsen MT, Gilbert MTP (2014) Global distribution of chelonid fibropapilloma-associated herpesvirus among clinically healthy sea turtles. *BMC Evol Biol* 14:206
- Balazs GH (1991) Current status of fibropapillomatosis in the Hawaiian green turtle, *Chelonia mydas*. In: Balazs GH, Pooley SG (eds) Research plan for marine turtle fibropa-

- pilloma. NOAA Tech Memo NMFS-SWFSC-156. US Department of Commerce, Honolulu, HI, p 47–57
- Baptistotte C (2007) Caracterização espacial e temporal da fibropapilomatose em tartarugas marinhas da costa brasileira. PhD thesis, Universidade de São Paulo, Piracicaba
- Baptistotte C (2016) Fibropapillomatosis in sea turtles from South America—Brazil, Uruguay, and Argentina. In: Hargrove S, Work T, Brunson S, Foley AM, Balazs G (eds) Proc 2015 Int summit on fibropapillomatosis: global status, trends and population impacts. NOAA Tech Memo NMFS-PIFSC-54. US Department of Commerce, Honolulu, HI, p 22–25
- ✦ Bastos KV, Machado LP, Joyeux JC, Ferreira JS, Militão FP, Fernandes VO, Santos RG (2022) Coastal degradation impacts on green turtle's (*Chelonia mydas*) diet in south-eastern Brazil: nutritional richness and health. *Sci Total Environ* 823:153593
- Bennett P, Keuper-Bennett U, Balazs G (2000) Photographic evidence for the regression of fibropapillomas afflicting green turtles at Honokawai, Maui, in the Hawaiian Islands. In: Kalb H, Wibbels T (eds) Proc 19th Ann Symp sea turtle biology and conservation, 2–6 March 1999, South Padre Island, TX. NOAA Tech Memo NMFS-SEFSC-443. National Marine Fisheries Service, Miami, FL, p 37–39
- Bjorndal KA (1996) Foraging ecology and nutrition of sea turtles. In: Lutz PL, Musick JA (eds) The biology of sea turtles. CRC Press, Boca Raton, FL, p 199–231
- ✦ Bjorndal KA, Bolten AB, Chaloupka MY (2000) Green turtle somatic growth model: evidence for density dependence. *Ecol Appl* 10:269–282
- Bolten AB (1999) Techniques for measuring sea turtles. In: Eckert KL, Bjorndal KA, Abreu-Grobois FA, Donnelly M (eds) Research and management techniques for the conservation of sea turtles. Publication No. 4. IUCN/SSC Marine Turtle Specialist Group, Washington, DC, p 110–114
- ✦ Broderick A, Patricio A (2019) *Chelonia mydas* (South Atlantic subpopulation). IUCN Red List of Threatened Species 2019: e.T142121866A142086337. <https://dx.doi.org/10.2305/IUCN.UK.2019-2.RLTS.T142121866A142086337.en> (accessed 28 October 2023)
- ✦ Carpentier AS, Jean C, Barret M, Chassagneux A, Ciccione S (2016) Stability of facial scale patterns on green sea turtles *Chelonia mydas* over time: a validation for the use of a photo-identification method. *J Exp Mar Biol Ecol* 476: 15–21
- Centro TAMAR/ICMBio (2017) Manual para marcação e biometria de tartarugas marinhas. Ministério do Meio Ambiente, Instituto Chico Mendes de Conservação da Biodiversidade (ICMBio), Centro Nacional de Pesquisa e Conservação de Tartarugas Marinhas e Fundação Pró-Tamar, Brasília
- ✦ Chaloupka M, Limpus C (2005) Estimates of sex- and age-class-specific survival probabilities for a southern Great Barrier Reef green sea turtle population. *Mar Biol* 146: 1251–1261
- ✦ Chaloupka M, Balazs GH, Work TM (2009) Rise and fall over 26 years of a marine epizootic in Hawaiian green sea turtles. *J Wildl Dis* 45:1138–1142
- ✦ Cray C, Varella R, Bossart GD, Lutz P (2001) Altered in vitro immune responses in green turtles (*Chelonia mydas*) with fibropapillomatosis. *J Zoo Wildl Med* 32:436–440
- Cremer MJ (2006) O estuário da Baía da Babitonga. In: Cremer MJ, Morales PRD, Oliveira TMN (eds) Diagnóstico ambiental da Baía da Babitonga. UNIVILLE, Joinville, p 15–19
- Cremer MJ, Sartori CM (2009) Tartarugas marinhas no litoral norte de Santa Catarina. *Rev UNIVILLE* 14:57–63
- ✦ Cremer MJ, Souza TF, Domiciano IG, Goldberg DW, Wanderlinde J (2020) Tartarugas marinhas no litoral norte de Santa Catarina e Baía Babitonga. *Rev CEPsul Biodivers Conserv Mar* 9:eb2020002
- ✦ Domiciano IG, Domit C, Bracarense APFRL (2017) The green turtle *Chelonia mydas* as a marine and coastal environmental sentinels: anthropogenic activities and diseases. *Semin Cienc Agrar* 38:3417–3434
- ✦ Domiciano IG, Broadhurst MK, Domit C, Flaiban KKM, Goldberg DW, Fritzen JTT, Bracarense APFRL (2019) Chelonid alphaherpesvirus 5 DNA in fibropapillomatosis-affected *Chelonia mydas*. *EcoHealth* 16:248–259
- Ehrhart LM, Ogren LH (1999) Studies in foraging habits: capturing and handling turtles. In: Eckert KL, Bjorndal KA, Abreu-Grobois FA, Donnelly M (eds) Research and management techniques for the conservation of sea turtles. Publication No. 4. IUCN/SSC Marine Turtle Specialist Group, Washington, DC, p 61–64
- ✦ Ene A, Su M, Lemaire S, Rose C and others (2005) Distribution of chelonid fibropapillomatosis-associated herpesvirus variants in Florida: molecular genetic evidence for infection of turtles following recruitment to neritic developmental habitats. *J Wildl Dis* 41:489–497
- ✦ Espinoza-Rodríguez N, Barrios-Garrido H (2021) Fibropapillomatosis in immature Green Turtle (*Chelonia mydas*) from the Gulf of Venezuela. *Rev Cient (Maracaibo)* XXXII:1–6
- ✦ Farrell JA, Yetsko K, Whitmore L, Whilde J and others (2021) Environmental DNA monitoring of oncogenic viral shedding and genomic profiling of sea turtle fibropapillomatosis reveals unusual viral dynamics. *Commun Biol* 4:565
- ✦ Farrell JA, Whitmore L, Mashkour N, Rollinson Ramia DR and others (2022) Detection and population genomics of sea turtle species via noninvasive environmental DNA analysis of nesting beach sand tracks and oceanic water. *Mol Ecol Resour* 22:2471–2493
- ✦ Flint M (2013) Free-ranging sea turtle health. In: Wyneken J, Lohmann KJ, Musick JA (eds) The biology of sea turtles, Vol 3. CRC Press, Boca Raton, FL, p 379–397
- ✦ Foley AM, Schroeder BA, Redlow AE, Fick-Child KJ, Teas WG (2005) Fibropapillomatosis in stranded green turtles (*Chelonia mydas*) from the eastern United States (1980–98): trends and associations with environmental factors. *J Wildl Dis* 41:29–41
- ✦ Fox J, Marquez M (2023) RcmdrMisc: R commander miscellaneous functions. R package version 2.9-1. <https://CRAN.R-project.org/package=RcmdrMisc>
- ✦ Gattamorta MA, Gavilan SA, Silva FJL, Zamana RR and others (2022) First report of fibropapillomatosis (FP) and *Chelonid alphaherpesvirus 5* (ChHV5) in a green sea turtle (*Chelonia mydas*) from the historically fibropapillomatosis-free Fernando de Noronha Archipelago, Northeastern Brazil. *Braz J Vet Res Anim Sci* 59:e181776
- George RH (1996) Health problems and diseases of sea turtles. In: Lutz PL, Musick JA (eds) The biology of sea turtles. CRC Press, Boca Raton, FL, p 363–386
- ✦ Gerhardinger LC, Herbst DF, Carvalho FG, Freitas RR and others (2021) Diagnóstico socioambiental do Ecossistema Babitonga. *Rev CEPsul Biodivers Conserv Mar* 10: e2021002
- ✦ Glickman ME, Rao SR, Schultz MR (2014) False discovery rate control is a recommended alternative to Bonferroni-type adjustments in health studies. *J Clin Epidemiol* 67: 850–857

- Hamann M, Godfrey MH, Seminoff JA, Arthur K and others (2010) Global research priorities for sea turtles: informing management and conservation in the 21st century. *Endang Species Res* 11:245–269
- Hancock JM, Choma J, Mainye L, Wambi P and others (2023) Using photo-ID to document and monitor the prevalence of fibropapilloma tumours in a foraging aggregation of green turtles. *Front Mar Sci* 10:1217683
- Herbst LH (1994) Fibropapillomatosis of marine turtles. *Annu Rev Fish Dis* 4:389–425
- Herbst LH, Klein PA (1995) Green turtle fibropapillomatosis: challenges to assessing the role of environmental cofactors. *Environ Health Perspect* 103:27
- Herbst L, Ene A, Su M, Desalle R, Lenz J (2004) Tumor outbreaks in marine turtles are not due to recent herpesvirus mutations. *Curr Biol* 14:R697–R699
- IBAMA (Instituto Brasileiro do Meio Ambiente e dos Recursos Naturais Renováveis) (2014) Portaria n° 444, de 17 de dezembro de 2014. Reconhecer como espécies da fauna brasileira ameaçadas de extinção aquelas constantes da 'Lista Nacional Oficial de Espécies da Fauna Ameaçadas de Extinção'. *Diário Oficial da União: seção 1, Brasília, DF, n. 245, p. 121, 18 dez. 2014*
- IBAMA (2022) Portaria n° 148, de 7 de junho de 2022. Altera os Anexos da Portaria n° 443, de 17 de dezembro de 2014, da Portaria n° 444, de 17 de dezembro de 2014, e da Portaria n° 445, de 17 de dezembro de 2014, referentes à atualização da Lista Nacional de Espécies Ameaçadas de Extinção. *Diário Oficial da União: seção 1, Brasília, DF, n. 108, p. 74, 8 jun. 2022*
- Jones K, Ariel E, Burgess G, Read M (2016) A review of fibropapillomatosis in green turtles (*Chelonia mydas*). *Vet J* 212:48–57
- Jones K, Burgess G, Budd AM, Huerlimann R, Mashkour N, Ariel E (2020) Molecular evidence for horizontal transmission of chelonid alphaherpesvirus 5 at green turtle (*Chelonia mydas*) foraging grounds in Queensland, Australia. *PLOS ONE* 15:e0227268
- Kassambara A (2023) rstatix: pipe-friendly framework for basic statistical tests. R package version 0.7.2. <https://CRAN.R-project.org/package=rstatix>
- Kelley JR, Kelley KL, Savage AE, Mansfield KL (2022) Novel disease state model finds most juvenile green turtles develop and recover from fibropapillomatosis. *Ecosphere* 13:e4000
- Lackovich JK, Brown DR, Homer BL, Garber RL and others (1999) Association of herpesvirus with fibropapillomatosis of the green turtle *Chelonia mydas* and the loggerhead turtle *Caretta caretta* in Florida. *Dis Aquat Org* 37:89–97
- Lenz AJ, Avens L, Borges-Martins M (2017) Age and growth of juvenile green turtles *Chelonia mydas* in the western South Atlantic Ocean. *Mar Ecol Prog Ser* 568:191–201
- Machado Guimarães S, Mas Gitirana H, Vidal Wanderley A, Monteiro-Neto C, Lobo-Hajdu G (2013) Evidence of regression of fibropapillomas in juvenile green turtles *Chelonia mydas* caught in Niterói, southeast Brazil. *Dis Aquat Org* 102:243–247
- Mazaris AD, Schofield G, Gkazinou C, Almpandou V, Hays GC (2017) Global sea turtle conservation successes. *Sci Adv* 3:e1600730
- Miguel C, Costa PG, Bianchini A, Luzardo OLP, Vianna MRM, Santos MRD (2022) Health condition of *Chelonia mydas* from a foraging area affected by the tailings of a collapsed dam in southeast Brazil. *Sci Total Environ* 821:153353
- MMA/SBio/DCME (Ministério do Meio Ambiente/Secretaria de Biodiversidade/Departamento de Conservação e Manejo de Espécies) (2023) Áreas prioritárias para a Conservação, Utilização Sustentável e Repartição dos Benefícios da Biodiversidade 2ª Atualização. Série Biodiversidade 56. MMA/SBio/DCME, Brasília
- Monteiro J, Duarte M, Amadou K, Barbosa C and others (2021) Fibropapillomatosis and the chelonid alphaherpesvirus 5 in green turtles from West Africa. *EcoHealth* 18:229–240
- Morrison CL, Iwanowicz L, Work T, Fahsbender E and others (2018) Genomic evolution, recombination, and inter-strain diversity of chelonid alphaherpesvirus 5 from Florida and Hawaii green sea turtles with fibropapillomatosis. *PeerJ* 6:e4386
- Muñoz Tenería FA, Labrada-Martagón V, Herrera-Pavón RL, Work TM, González-Ballesteros E, Negrete-Philippe AC, Maldonado-Saldaña G (2022) Fibropapillomatosis dynamics in green sea turtles *Chelonia mydas* over 15 years of monitoring in Akumal Bay, Quintana Roo, Mexico. *Dis Aquat Org* 149:133–143
- Neves-Ferreira I, Mello-Fonseca J, Ferreira CEL (2023) Photo-identification shows the spatio-temporal distribution of two sea turtle species in a Brazilian developmental foraging ground. *Mar Biol* 170:83
- Page-Karjian A, Torres F, Zhang J, Rivera S and others (2012) Presence of chelonid fibropapilloma-associated herpesvirus in tumored and non-tumored green turtles, as detected by polymerase chain reaction, in endemic and non-endemic aggregations, Puerto Rico. *SpringerPlus* 1:35
- Page-Karjian A, Stacy NI, Morgan AN, Coppentrath CM, Manire CA, Herbst LH, Perrault JR (2022) Morphologic and physiologic characteristics of green sea turtle (*Chelonia mydas*) hatchlings in southeastern Florida, USA. *J Comp Physiol B* 192:751–764
- Patrício AR, Herbst LH, Duarte A, Vélez-Zuazo X and others (2012) Global phylogeography and evolution of chelonid fibropapilloma-associated herpesvirus. *J Gen Virol* 93:1035–1045
- Patrício AR, Diez CE, van Dam RP, Godley BJ (2016) Novel insights into the dynamics of green turtle fibropapillomatosis. *Mar Ecol Prog Ser* 547:247–255
- Pérez YAA, Lima SR, Martinez-Souza G, Gião T and others (2024) First case report of fibropapillomatosis tumor regression identified through photoidentification and histopathology in a *Chelonia mydas* in Itapirubá, Santa Catarina, Brazil. *Open Vet J* 14:3649
- Quackenbush SL, Casey RN, Murcek RJ, Paul TA and others (2001) Quantitative analysis of herpesvirus sequences from normal tissue and fibropapillomas of marine turtles with real-time PCR. *Virology* 287:105–111
- Core Team (2023) R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna
- Rees AF, Alfaro-Shigueto J, Barata PCR, Bjorndal KA and others (2016) Are we working towards global research priorities for management and conservation of sea turtles? *Endang Species Res* 31:337–382
- Rodenbusch CR, Baptistotte C, Werneck MR, Pires TT and others (2014) Fibropapillomatosis in green turtles *Chelonia mydas* in Brazil: characteristics of tumors and virus. *Dis Aquat Org* 111:207–217
- Roost T, Schies JA, Girondot M, Robin JP and others (2022) Fibropapillomatosis prevalence and distribution in immature green turtles (*Chelonia mydas*) in Martinique Island (Lesser Antilles). *EcoHealth* 19:190–202

- ✦ Rossi S, Sánchez-Sarmiento AM, Vanstreels RET, Santos RG and others (2016) Challenges in evaluating the severity of fibropapillomatosis: a proposal for objective index and score system for green sea turtles (*Chelonia mydas*) in Brazil. PLOS ONE 11:e0167632
- ✦ Rossi S, Sánchez-Sarmiento AM, Santos RG, Zamana RR and others (2019) Monitoring green sea turtles in Brazilian feeding areas: relating body condition index to fibropapillomatosis prevalence. J Mar Biol Assoc UK 99: 1879–1887
- ✦ Santos RG, Martins AS, Torezani E, Baptistotte C and others (2010) Relationship between fibropapillomatosis and environmental quality: a case study with *Chelonia mydas* off Brazil. Dis Aquat Org 89:87–95
- ✦ Santos MRD, Martins AS, Baptistotte C, Work TM (2015) Health condition of juvenile *Chelonia mydas* related to fibropapillomatosis in southeast Brazil. Dis Aquat Org 115:193–201
- ✦ Santos MRD, Ferreira PD Jr, Nóbrega YC, Merçon J, Pereira TM, Gomes LC (2017) Stress response of juvenile green sea turtles (*Chelonia mydas*) with different fibropapillomatosis scores. J Wildl Dis 53:653–656
- ✦ Seminoff JA (2004) *Chelonia mydas*. The IUCN Red List of Threatened Species 2004: e.T4615A11037468. <https://dx.doi.org/10.2305/IUCN.UK.2004.RLTS.T4615A11037468.en> (accessed 28 October 2023)
- ✦ Silva-Júnior ES, Farias DSD, Bomfim AC, Freire ACB and others (2019) Stranded marine turtles in Northeastern Brazil: Incidence and spatial–temporal distribution of fibropapillomatosis. Chelonian Conserv Biol 18:249–258
- ✦ Souza TF (2016) Ecologia alimentar da tartaruga-verde, *Chelonia mydas*, no litoral norte de Santa Catarina. MSc dissertation, Universidade da Região de Joinville
- ✦ Tagliolatto AB, Guimarães SM, Lobo-Hajdu G, Monteiro-Neto C (2016) Characterization of fibropapillomatosis in green turtles *Chelonia mydas* (Cheloniidae) captured in a foraging area in southeastern Brazil. Dis Aquat Org 121: 233–240
- ✦ Van Houtan KS, Hargrove SK, Balazs GH (2010) Land use, macroalgae, and a tumor-forming disease in marine turtles. PLOS ONE 5:e12900
- ✦ Vanstreels RET, Durant A, Santos AP, Santos RG and others (2023) Exploring the relationship between environmental drivers and the manifestation of fibropapillomatosis in green turtles (*Chelonia mydas*) in eastern Brazil. PLOS ONE 18:e0290312
- ✦ Wickham H (2016) ggplot2: elegant graphics for data analysis. Springer-Verlag, New York, NY
- ✦ Wickham H, Bryan J (2023) readxl: read Excel files. R package version 1.4.2. <https://CRAN.R-project.org/package=readxl>
- ✦ Work TM, Balazs GH (1999) Relating tumor score to hematology in green turtles with fibropapillomatosis in Hawaii. J Wildl Dis 35:804–807
- ✦ Work TM, Rameyer RA, Balazs GH, Cray C, Chang SP (2001) Immune status of free-ranging green turtles with fibropapillomatosis from Hawaii. J Wildl Dis 37:574–581
- ✦ Work TM, Balazs GH, Rameyer RA, Morris RA (2004) Retrospective pathology survey of green turtles *Chelonia mydas* with fibropapillomatosis in the Hawaiian Islands, 1993–2003. Dis Aquat Org 62:163–176
- ✦ Work TM, Dagenais J, Balazs GH, Schettler N, Ackermann M (2015) Dynamics of virus shedding and *in situ* confirmation of chelonid herpesvirus 5 in Hawaiian green turtles with fibropapillomatosis. Vet Pathol 52:1195–1201
- ✦ Worm B, Barbier EB, Beaumont N, Duffy JE and others (2006) Impacts of biodiversity loss on ocean ecosystem services. Science 314:787–790
- ✦ Wyneken J, Lohmann KJ, Musick JA (2013) The biology of sea turtles, Vol 3. CRC Press, Boca Raton, FL
- ✦ Zamana RR, Gattamorta MA, Ochoa PFC, Navas-Suárez PE and others (2021) High occurrence of chelonid alphaherpesvirus 5 (ChHV5) in green sea turtles *Chelonia mydas* with and without fibropapillomatosis in feeding areas of the São Paulo Coast, Brazil. J Aquat Anim Health 33: 252–263

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